

1 ***Caenorhabditis elegans* Exhibits Positive Gravitaxis**

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9 10 **Abstract**

11 Whether or not the micro swimmer *Caenorhabditis elegans* senses and respond to gravity is
12 unknown. We find that *C. elegans* aligns its swimming direction with that of the gravity vector
13 (positive gravitaxis). When placed in an aqueous solution that is denser than the animals, they
14 still orient downwards, indicating that non-uniform mass distribution and/or hydrodynamic
15 effects are not responsible for animal's downward orientation. Paralyzed worms and worms
16 with globally disrupted sensory cilia do not change orientation as they settle in solution,
17 indicating that gravitaxis is an active behavior that requires gravisensation. Other types of
18 sensory driven orientation behaviors cannot explain our observed downward orientation. Like
19 other neural behaviors, the ability to respond to gravity declines with age. Our study establishes
20 gravitaxis in the micro swimmer *C. elegans* and suggests that *C. elegans* can be used as a
21 genetically tractable system to study molecular and neural mechanisms of gravity sensing and
22 orientation.

25 **Significance Statement**

26 Understanding how animals respond to gravity is not only of fundamental scientific interest,
27 but has clinical relevance, given the prevalence of postural instability in aged individuals.
28 Determining whether *C. elegans* responds to gravity is important for mechanistic studies of
29 gravity sensing in an experimentally tractable animal, for a better understanding of nematode
30 ecology and evolution, and for studying biological effects of microgravity. Our experiments,
31 which indicate that *C. elegans* senses and responds to gravity, set the stage for mechanistic
32 studies on molecular mechanisms of gravity sensing.

33

34 **Introduction**

35 Gravity plays an important role in most life forms on earth, ranging from single cells to
36 plants and animals. Plants' roots grow in the direction of gravity (positive gravitropism) and
37 shoots grow in the opposite direction (negative gravitropism) to optimize nutrient uptake and
38 exposure to light (1). Aquatic invertebrates use gravity cues to help navigate in the vertical
39 dimension (2). Both terrestrial and aquatic vertebrates know which direction is up. While there
40 are gravity sensory organ differences that relate to the unique ecologies across phylogeny, there
41 are also similarities in the anatomical and physiological principles of such organs. Despite the
42 importance of gravity sensing to life on earth, many molecular components of sensing and
43 responding to gravity remain unknown.

44 In this study, we examine whether the nematode *Caenorhabditis elegans* (*C. elegans*)
45 senses and responds to the direction of gravity. *C. elegans* offers important experimental
46 advantages, including a small and simple nervous system, accessibility to rapid genetic
47 manipulation and to other powerful experimental tools, and ease of cultivation. There is only
48 one report suggesting that *C. elegans* suspended in solution may orient with the gravitational
49 field (3). Since *C. elegans* is heavier than suspending buffers typically used in laboratories, it

50 settles when suspended in solution (4). Our observations suggest that as wild type animals settle,
51 they also orient their direction of swimming to align with the direction of the gravity vector.
52 Does *C. elegans* sense gravity? Is its response to gravitational forces passive or active? We set
53 to answer these questions in this study.

54

55 **Materials and methods:**

56 **Worm preparation:** On the day prior to the experiment, well-fed fourth larval stage
57 hermaphrodites were placed on an agar plate containing a bacterial lawn of OP50. Day-one
58 adult worms were harvested from the agar plate by floating the worms in M9 buffer and then
59 transferring the worm suspension into a 1.5 mL conical tube. Following centrifugation (4000-
60 5000 rpm) for a few seconds to sediment the worms, the supernatant was decanted. The worms
61 were then washed three times with 1 mL M9 buffer by repeating the centrifugation/decanting
62 steps. We experimented mostly with “recently-fed” worms - the time elapsed from floating the
63 worms off their cultivation plate to the completion of the experiment was < 30 min. A few of
64 the experiments were carried out with “starved” worms - the time elapsed from floating the
65 worms off their cultivation plate to the start of the experiment was ~ 1 hour.

66 To paralyze wild-type worms, we suspended the worms in one milliliter M9 buffer in a
67 1.5 mL conical tube and placed them for one hour in a water bath at 40°C. The experiment was
68 then performed at room temperature (21~22°C) within 30 minutes from the removal of the
69 animals from the water bath. Observations of these worms showed that they were fully
70 paralyzed for over 30 minutes after the heat shock

71 **High density buffer:** To achieve a density greater than that of the worms, we mixed a colloidal
72 silica solution (LUDOX HS-40, Sigma, density: 1.3 g/mL at 25°C (5) with M9 buffer at a
73 volume ratio 1:2 to form a solution with density of 1.1 g/mL, which is slightly greater than the

74 worm's density (~ 1.07 g/mL (6)). The mixture density was measured directly by weighing one
75 ml of solution. At the density used in our experiment, the suspension behaves like a Newtonian
76 liquid with a viscosity approximately 7 times that of water (7).

77 **Experimental Apparatus:** A cuboid polystyrene cuvette with a square cross-section $12\text{ mm} \times$
78 12 mm and heights ranging from 45 to 200 mm filled with M9 buffer at room temperature
79 ($21\sim 22^\circ\text{C}$) were used in our settling experiments. $20\ \mu\text{L}$ of a worm suspension at a
80 concentration of about 1.5 worms per microliter was extracted with a plastic tip pipette and
81 transferred into the cuvette by slowly expelling the worms into the cuvette solution either above
82 or just below the liquid surface.

83 **Imaging:** The worms were monitored with two cameras (IMAGINGSOURCE DMK 33GP031
84 with a 25 mm lens and IMAGINGSOURCE DMK 22BUC03 with a 12 mm lens) acquiring
85 images at 30 frames per second from two orthogonal planes (**Fig. 1**). One camera focused on
86 the X-Z plane and the other on the Y-Z plane at the cuvette's center. Each image size was 640
87 $\times 480$ pixels, which results in an aspect ratio of 4:3. As it settled, a worm stayed about 10 s
88 within the field of view of the two cameras, resulting in about 300 double frames for each
89 worm. Images were processed with a Matlab R2018b graphical user interface (GUI), following
90 the image processing scheme described in (8) and outlined in the Supporting Information (SI-
91 Section S1).

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97 Results

98 Wild Type (WT) *C. elegans* young adults align their swimming direction with the direction 99 of gravity

100 We inserted first day adult, recently-fed, wild-type (WT) animals just beneath the water
101 surface at the top of our cuvette and monitored the animals' orientation (θ , φ) as a function of
102 time (**Fig. 1**). Here, θ and φ are, respectively, the polar (inclination) and azimuthal angles.
103 $\theta = 180^\circ$ is the direction of gravity. Since the worms (density ~ 1.07 g/mL) are heavier than
104 water (~ 1 g/mL), they settled to the cuvette's bottom.

105 As time went by, the WT worms varied their swimming direction to align with the
106 direction of the gravity vector. **Fig. 2** exemplifies this behavior. The figure depicts time-lapsed
107 frames, 1-second apart, of a WT young adult animal, inserted beneath the water surface and
108 settling in our cuvette. In the first image (A), the animal is a distance ~ 6.5 mm beneath the
109 water surface and faces nearly upwards $\theta \sim 5^\circ$. As time increases, the polar angle θ gradually
110 increases. In the last frame (J), the animal is ~ 11.5 mm beneath the water surface and its polar
111 angle $\theta \sim 142^\circ$. The worm has changed its direction of swimming from nearly upwards to
112 nearly downwards. This behavior is exhibited more clearly in panel K, wherein we translated
113 the skeletons of the animal to position their centroids at the same point. **Fig. 2K** illustrates the
114 animal's tendency to rotate to align its direction of swimming with the direction of gravity.

115 Regardless of initial orientation, given enough time, the animals oriented themselves in
116 the direction of gravity independent of their azimuthal position (SI-Section S2). **Fig. 3** depicts
117 the kernel density estimate KDE $f(\theta)$ (an approximation of the probability distribution function,
118 *pdf*) (9) of animals' orientations at various depths beneath the liquid surface. Close to the
119 liquid's surface (shortly after release), $f(\theta)$ is nearly symmetric about the horizontal direction
120 ($\theta=90^\circ$), indicating lack of orientation bias and equal probability towards upward and
121 downward swimming. The KDE resembles a *sin* function that corresponds to a uniform *pdf* in

122 the θ -direction. As time goes by, the KDE function skews in the direction of increasing polar
123 angles, indicating that as the worms descend, they rotate to increase their polar angle and align
124 their direction of swimming with the direction of gravity. Our KDE resembles the von-Mises
125 Fisher directional pdf (10) with the mean inclination (polar) angle $\theta = 180^\circ$ (SI-Section S4):

$$126 \quad f(\theta, \varphi) = \frac{\lambda}{4\pi \text{Sinh}\lambda} e^{\lambda \cos(\pi - \theta)}, \quad (1)$$

127 where the *concentration parameter* λ (reciprocal measure of dispersion) is analogous to the
128 inverse of the variance in a normal distribution. $\lambda \rightarrow 0$ corresponds to a uniform distribution.
129 Since our data is independent of the azimuthal angle φ , we integrate in φ to obtain

$$130 \quad f(\theta) = \frac{\lambda}{2\text{Sinh}\lambda} e^{\lambda \cos(\pi - \theta)} \sin(\pi - \theta). \quad (2)$$

131 When $\lambda \geq 1$ and $\lambda \geq 3$, over 73% and 95% of the animals are oriented, respectively, at a polar
132 angle $\theta > 90^\circ$. We compute the concentration parameter λ for our data by fitting the cumulative
133 distribution function (cdf) associated with equation 2 (SI-section S4) to the experimental one.
134 When the animal is at depth $d = 4$ mm beneath the surface $\lambda \sim 0.2$ (nearly uniform distribution).
135 As the animal's depth increases (the animal has more time to align with the direction of gravity),
136 the skewness of the KDE and the magnitude of λ increase as well. For the well-fed WT animals
137 λ increases at the approximate rate of 0.07 per mm of depth until it asymptotes to ~ 4.3 at ~ 60
138 mm, and approximately retains this value at depths exceeding 60 mm. KDEs at depths 120 mm
139 $< d < 200$ mm nearly overlap (SI – Section S5). The inset in **Fig. 3** depicts the concentration
140 parameter λ as a function of the animal's depth (d , mm) beneath the liquid surface. The data is
141 correlated with the expression

$$142 \quad \lambda(d) = \lambda_\infty (1 - e^{-\beta d}), \quad (3)$$

143 where $\lambda_\infty \sim 4.36$ and $\beta \sim 0.03$ mm⁻¹. Equation (3) illustrates that after the worm reaches a certain
144 depth, the orientation of the animals attains a stationary state.

145 The sedimentation velocity of a rigid, cylindrical rod depends on the rod's orientation
146 with respect to its direction of motion (11); rigid rods settle faster when aligned broadside than

147 when their axis parallels the direction of motion. To test whether this applies to *C. elegans* and
148 to approximately correlate the animal's depth with its residence time in solution, we examined
149 the worm's translational and angular velocities. **Fig. 4** depicts the velocity (U) of young adult
150 WT worms' centroid in the direction of swimming as a function of $(-\cos \theta)$. The data is
151 scattered along a straight line and correlates well ($R^2=0.89$, solid line) with the expression.

$$152 \quad U = U_s - U_g \cos \theta. \quad (4)$$

153 We interpret U_s as the animal's swimming velocity and U_g as the sedimentation velocity in
154 the gravitational field. The term $(-U_g \cos \theta)$ is the projection of the sedimentation velocity on
155 the animal's swimming direction. In contrast to rigid cylindrical rods, the worm's settling
156 velocity U_g depends only weakly on orientation (θ). This perhaps results from the worm not
157 being perfectly straight and rigid. We estimate $\overline{U_s} \approx 411 \mu\text{m/s}$ and $\overline{U_g} \approx 432 \mu\text{m/s}$. The
158 angular velocity $\omega = \frac{d\theta}{dt}$ varied widely, but never exceeded 28 degrees/s .

159 To examine reproducibility of our data, we repeated our experiments in two continents
160 (USA and Taiwan) and a few weeks apart and obtained similar results (e.g., **SI - Fig. S3**). We
161 also demonstrated that the animals' azimuthal angle φ was uniformly distributed (**SI - Fig. S4**),
162 suggesting that convective currents in our apparatus, if any, are unlikely to have biased our data.

163 In summary, WT animals align their swimming direction with the gravity vector. Can this
164 alignment be attributed to non-uniform mass distribution along the animal's length?

165

166 **Paralyzed WT animals do not align with the direction of the gravity vector**

167 A plausible cause for animals to align with the direction of gravity is a non-uniform mass
168 distribution along the animal's body. Animals store fat primarily in the intestine (12), which is
169 not present in the anterior 1/5th of the worm's body; therefore, the worm may be head-heavy.

170 If significant, a non-uniform mass distribution would cause animals to rotate in a gravitational
171 field. To isolate potential effects of non-uniform mass distribution, we experimented with
172 paralyzed animals.

173 We achieved muscle paralysis by either exposing WT animals to high temperature (heat-
174 shock) or by testing animals that carry a mutation in the major muscle myosin gene *unc-54*
175 (13). Both heat-shocked WT animals and *unc-54* mutants maintained their initial inclination
176 (polar) angle and did not align with the direction of gravity as they descended. Their settling
177 velocity ($U_g \sim 432 \mu\text{m/s}$) is in excellent agreement with the estimated contribution of
178 gravitational settling to the velocity of WT animals (equation 3 and **Fig. 4**).

179 Both paralyzed WT and *unc-54* did not show any orientation preference during
180 sedimentation. The KDEs of the heat-shocked WT (**SI-Fig. S10**, $4 \text{ mm} < d < 100 \text{ mm}$, and **Fig.**
181 **5**, $120 \text{ mm} < d < 200 \text{ mm}$) and *unc-54* (**Fig. S11**, $d = 40 \text{ mm}$, which is sufficiently far beneath
182 the water surface to allow animals to begin to adjust their polar angle) resemble a uniform
183 distribution in the polar angle and their descent angle θ is nearly symmetric with respect to
184 $\theta \sim 90^\circ$. Likewise, λ of paralyzed WT animals ranged from 0.01 to 0.33 consistent with a nearly
185 uniform distribution (inset in **Fig. 5**). Contrast **Figs. 5** and **S10** with **Figs. 3** and **S9**. The
186 difference is striking. Active WT animals rotate to align with the gravity vector while paralyzed
187 animals do not vary their polar angle θ during their descent. The kernel distribution estimates
188 of **Figs. 5** and **S10** are statistically distinct from the kernel distribution estimate of active
189 animals **Fig. 3** and **S9** ($p < 0.0001$, Mann Whitney test (9)).

190 In summary, the marked difference between active and paralyzed animals indicates that
191 the propensity to align with the gravity vector is mediated by active mechanisms. These
192 experiments lend further support to our earlier conclusion that factors such as convective
193 currents in our cuvette, if any, are not responsible for animal's orientation since they would

194 have similarly impacted active and inactive animals. Does the level of animal's activity affect
195 how fast it orients itself with the gravitational field?

196

197 **Starved animals and animals defective in muscle function (*unc-29*) align with the direction**
198 **of the gravity vector at a slower rate than well-fed WT animals.**

199 To determine whether the propensity for positive gravitaxis behavior is affected by the
200 dietary history or by mild impairment (non-paralysis) in body movements, we experimented
201 with starved (> 1 hour from last feeding) WT animals (**Figs. S13 and S14**) and with *unc-29*
202 mutants, which are mildly defective in muscle function due to a mutation in an acetylcholine
203 receptor subunit (14) (**SI Figs. S15 and S16**). In both cases, as the animals' depth beneath the
204 liquid surface (and residence time) increased, so did the skewness of their KDEs and the
205 magnitude of their concentration factor λ (**SI Fig. S17**), indicating that these animals still align
206 with the direction of the gravity vector; albeit at a slower rate than the well-fed, WT animals.
207 The concentration factor λ of the starved WT animals increased at the approximate rate of 0.03
208 mm^{-1} with depth, about half that of the well-fed animals, until it attained the nearly stationary
209 value of 3.2 at 100 mm and greater depths. The concentration factor λ of *unc-29* mutants
210 increased at the approximate rate of 0.007 mm^{-1} and attained $\lambda \sim 1.5$ at the depth of 200 mm,
211 which is the largest depth available in our experimental apparatus. Although the concentration
212 factor λ of *unc-29* mutants did not show fully saturation, it did increase with time, in clear
213 contrast to fully paralyzed mutants.

214 In summary, the rate of animal's alignment with the direction of the gravitational field
215 declines as the animal's swimming vigor decreases. Importantly, even animals with reduced
216 swimming vigor, when given enough time, orient to align with the gravity vector. Our
217 observations suggest that positive gravitaxis behavior does not solely rely on vigorous muscle
218 movements. Can the propensity to align with the direction of gravity be attributed to

219 hydrodynamic effects?

220

221 **Gravitaxis does not result from hydrodynamic effects**

222 We reasoned that if the downward swimming orientation were the result of interactions
223 between the flow field induced by the swimmer and the flow field associated with downward
224 sedimentation than, based on symmetry arguments, animals sedimenting *upward* should align
225 with the direction that is opposite to the direction to the gravity vector. To test this hypothesis,
226 we suspended well-fed WT animals beneath the surface of a LUDOX suspension that has
227 density slightly greater than that of the animals. Our experiments were complicated by the
228 animals floating to the surface and the LUDOX suspension having viscosity greater than water,
229 decreasing the rotational velocity of the animals and allowing them less time to orient in the
230 gravitational field. In contrast to the predictions based on hydrodynamic symmetry
231 considerations, the animals rotated to orient *downwards* and swim in the direction of the gravity
232 vector. **Fig. 6** (A-J) shows 10 video frames, spaced 1s apart, of a young adult, well-fed, WT
233 worm. The red dot indicates the position of the worm's head. In the 10-seconds period of
234 observation, the polar angle increased from an initial value of 49.1° in frame A to a value of
235 138.9° in frame J. Frame K depicts the skeletons of the worms from frames (A-J) shifted to
236 align their geometric centers.

237 **Fig. 7** depicts the kernel (probability) density estimate of the polar angle θ shortly after
238 the animals' introduction into the suspension, 5 s later, and 10 s later. At short times, the KDE
239 resembles a *sin* function, characteristic of a uniform *pdf* in θ . As time passes, the peak of the
240 kernel density estimate shifts to larger values of the angle θ and the magnitude of the
241 concentration factor λ increases from nearly zero to 2.7. Therefore, animals suspended in a
242 liquid that is either lighter (**Fig. 3**) or heavier (**Fig. 7**) than themselves rotate to swim in the

243 direction of the gravity vector. In summary, our observation of downward swimming even
244 when sedimenting upwards indicates that hydrodynamics are not responsible for animals'
245 orientation. Gravitaxis is not caused by hydrodynamic effects.

246 There is another important conclusion that we can draw from this experiment. In both
247 vertebrates and some aquatic invertebrates, gravity is sensed via the interaction between a proof
248 mass (a mass denser than its surrounding) and sensory cilia of specialized cells internal to the
249 animal. In contrast, certain insects, such as strepsiptera species, use their head as a proof mass
250 to sense gravity. As the animal's head falls, the head brushes against hairs between the head
251 and thorax (2). By monitoring the direction of hair deformation, the animal senses the direction
252 of gravity. Our experiment indicates that the gravity sensing mechanisms in *C. elegans* are
253 interior to the animal's body, as is the case in vertebrates and some aquatic invertebrates and
254 are not affected by the direction of buoyancy.

255

256 **Magnetotaxis and other taxis are not responsible for the worm orientation in** 257 **gravitational field**

258 Vidal-Gadea et al (15) report that *C. elegans* orients to the earth's magnetic field during
259 vertical burrowing migrations. Well-fed adult worms of the N2 Bristol strain, which was
260 isolated in the Northern Hemisphere, migrated up, while starved N2 worms migrated down. In
261 contrast, well-fed adult worms of the AB1 Adelaide strain, which was isolated in the Southern
262 Hemisphere, migrated down while starved AB1 worms migrated up in response to the same
263 magnetic field. We have not observed similar tendencies in our experiments with *C. elegans* in
264 solution. In our experiments, both well-fed (**Figs. 2 and 3**) and starved (SI Figs. S13, S14, and
265 S17) WT animals oriented with the direction of the gravity field and swam downwards. Well-
266 fed AB1 worms (SI **Figs. S18 – S20**), like well-fed N2 worms, oriented downwards in the

267 vertical liquid column. Hence, the taxis mechanisms identified in reference (15) are unlikely to
268 explain our observations.

269 Since our experiments took place in a vessel subjected to uniform room light and
270 temperature, there is no gradient of light intensity or temperature and therefore no phototaxis
271 or thermotactic stimulation. While *C. elegans* prefers low oxygen tensions (16, 17), our
272 observations are unlikely to be explained by aerotaxis behavior because the concentrations of
273 gases are nearly uniform in our aqueous column, aside from O₂ consumption and CO₂
274 production by the worms, which is likely to be negligible on the time scale of our experiment.
275 Moreover, whereas in a low water density solution, the animals sedimented downward and
276 aggregated at the bottom of the water column, in high-density solution, the animals sedimented
277 upwards and congregated at the top of the water column. Any gaseous gradient caused by
278 consumption of oxygen or generation of carbon dioxide by aggregated animals would be
279 reversed in the high-density solution. Our observation that animal orientated downward
280 regardless of where the animals aggregated (at the top or the bottom) further indicates that
281 gaseous gradients do not explain our observations. In conclusion, given our exclusion of other
282 known taxis behaviors, our observations indicate that the animals respond to gravity when
283 swimming in an aqueous solution.

284

285 **Gravity sensing and ability to orient in gravitational field declines with age**

286 Many of *C. elegans* sensing capabilities deteriorate with age (18). Some of this
287 deterioration is identifiable with specific neuronal deficits. For example, the sensory dendrites
288 of the FLP and PVD neurons, which are required for response to certain mechanical stimuli,
289 degenerate with age (19). To examine aging worms' ability to sense and orient in gravitational
290 field, we measured the angle of descent as a function of animal's age (**Fig. 8**) when located 40

291 mm beneath the liquid surface. The aged animal (day 6) has a broader kernel density estimate
292 than the young adult (day 1), indicating that a greater fraction of the animals failed to align
293 with the direction of gravity. The concentration parameter λ (inset in **Fig. 8**) decreases as the
294 animal's age increases. Day 1 and day 2 adults animals behaved similarly with $\lambda = 2.9$ (N =
295 87) and 3.1 (N = 62), respectively. After day 2 of adulthood, there was a gradual decline in λ .
296 At day 6 of adulthood, which corresponds to a mid-life aged animal, $\lambda \sim 1.4$ (N = 50), which
297 is significantly smaller than that of day 1 adults ($p = 0.0012$, Mann Whitney test). In the age
298 range between 1 and 5 days, there is a gradual decline in the animals' ability to react to
299 gravitational field.

300

301 **Gravitaxis requires sensory neurons function**

302 We reasoned that if the worm senses gravity and deliberately orients in the downward
303 direction, we should be able to impair this behavior by selectively disrupting sensation with
304 minimal impairment of movement. Many sensory functions of *C. elegans* such as olfaction,
305 gustation, thermosensation, nose-touch, magnetoreception, and electrosensation are mediated
306 by neurons that extend cilia to the nose of the animal. We hypothesize that gravity sensation
307 too is mediated by ciliated sensory neurons. To test this hypothesis, we analyzed the angle of
308 descent of animals mutant for the gene *osm-6* or for the gene *che-2*, which encode, respectively,
309 intra flagellar protein 52 (IFT52) and IFT80 and in which sensory cilia are globally disrupted
310 (20).

311 The polar angles of the mutants *che-2* and *osm-6* did not vary as they descended. The
312 kernel-density estimate plots (**Fig. 9**, 40 mm beneath liquid surface) suggest that both *che-2*
313 and *osm-6* retain nearly uniform kernel density estimates with $\lambda = 0.1$ (N = 51) and 0.6 (N =
314 70), respectively. Their kernel density estimates were statistically distinct from the WT control
315 ($p < 0.0001$, Mann Whitney Test). Our data suggests that the cilia-mutant animals show little

316 preference in their angle of descent. We conclude that ciliated sensory neurons are necessary
317 for gravitaxis.

318

319 **Discussion**

320 Hydrodynamists characterize motion of objects in fluids based on the relative importance
321 of inertial and viscous forces, quantified by the Reynolds number $Re=U D/\nu$. Where U is the
322 object velocity, D is the object's characteristic length, and ν is the kinematic viscosity of the
323 suspending fluid. The combined swimming and settling velocity U of *C. elegans* is less than 1
324 mm/s. The characteristic length scale, e.g., the diameter of *C. elegans*, $D\sim 80\ \mu\text{m}$ and the
325 kinematic viscosity of water $\nu\sim 10^{-6}\ \text{m}^2/\text{s}$. In all our experiments, $Re<0.1$, viscous effects
326 dominate, and the equations of motion are linear (Stokes equation). When inertia effects are
327 negligible ($Re\rightarrow 0$), cylindrical objects with fore–aft symmetry released with an inclination
328 angle θ sediment, in an unbounded medium, with the angle of release (11). Objects with non-
329 uniform mass distribution turn to bring their center of mass beneath their centroid. If *C. elegans*
330 were head heavy, it would eventually descend at $\theta=180^\circ$ when $Re\rightarrow 0$. In the presence of weak
331 inertia ($Re>0$), a cylindrical object turns to attain horizontal (broadside, $\theta=90^\circ$) posture and
332 then settles horizontally with its center of mass descending in the direction of gravity (21).

333 In our experiments, paralyzed WT animals settling in water retained their initial orientation
334 (polar) angle and did not show any tendency to align with the direction of the gravity vector,
335 consistent with low Reynolds number hydrodynamic theory for cylindrical objects with fore–
336 aft symmetry. This suggests that non-uniform mass distribution, if any, along the animal's
337 length is insignificant. Additionally, our data indicates lack of convective currents in the
338 experimental apparatus that might have affected animals' orientation. Since motion-impaired
339 animals do not align with the direction of the gravity vector, such alignment requires active
340 mechanisms.

341 Prior studies indicate that *C. elegans* traits such as gait-synchronization (22), tendency to
342 swim against the flow (rheotaxis) (23, 24), and tendency to swim along surfaces (bordertaxis)
343 (25) are involuntary and can be explained by simple mechanics. Could an interaction between
344 the flow-field induced by the animal's swimming gait and the flow-field induced by the
345 animal's sedimentation cause the animal to rotate and align with the gravity field? If such an
346 alignment mechanism existed, one would conclude, based on symmetry arguments, that
347 animals suspended in a liquid denser than them would align in the opposite direction to that of
348 the gravity vector. Our experiments with WT animals in LUDOX solution that is denser than
349 the animals (**Figs. 6 and 7**) indicate that this is not the case. Hence, we exclude hydrodynamics
350 as a possible explanation for gravitaxis.

351 Prior workers (15) reported that well-fed N2 strain (isolated in the Northern Hemisphere)
352 and the well-fed AB1 strain (isolated in Southern Hemisphere) crawl in opposite directions in
353 response to the same magnetic field. Moreover, starved N2 worms crawl in the opposite
354 direction to that of well-fed N2 worms in a magnetic field. In contrast, we observed that both
355 N2 and AB1 worms suspended in solution, regardless of being well-fed or starved, oriented
356 downwards in our vertical liquid column, demonstrating that mechanisms identified in
357 reference (15) do not explain our observations.

358 Our observations that *che-2* and *osm-6* mutants, which have the necessary motility to align
359 with the direction of gravity, fail to do so, further support our conclusion that gravitaxis in *C.*
360 *elegans* is deliberate, resulting from the animal's ability to sense the direction of gravity and
361 act on this information.

362 Gravity-sensing organs in invertebrates may be either external or internal to the animal's
363 body. The organ responsible for gravity sensing in *C. elegans* is still elusive. While a proof
364 mass similar to that seen in vertebrate inner ears has not been reported in ultrastructural studies

365 of *C. elegans*, it is possible that such a mass may have escaped detection due to its destruction
366 during tissue fixation and preparation processes.

367 WT *C. elegans* exhibits positive gravitaxis in both suspending medium that is lighter and a
368 suspending medium that is denser than the animal, demonstrating that gravity perception is not
369 affected by the density of the medium external to the animal. Therefore, the organ responsible
370 for graviatxis in *C. elegans* must be internal.

371 The evolutionary causes of positive gravitaxis behavior in *C. elegans* are a subject of
372 speculation. One possibility is that when dwelling in wet soil, downward migration would keep
373 the worms moist and away from the drying surface as well as distance them from air-borne
374 predators. Downward migration in bodies of water, may provide protection from predators as
375 well as direct the worms towards sources of food such as underwater flora and associated
376 bacteria.

377 Regardless of the reason for gravitaxis, we have here shown that the microscopic
378 nematode *C. elegans* orients its swimming direction to align with the direction of the gravity
379 vector, and that this behavior is not the result of an unequal distribution of mass, hydrodynamic
380 interactions, experimental artifacts, and other types of sensory-driven movements, or
381 hydrodynamic interactions. Taken together, our results indicate that *C. elegans* can sense and
382 respond to the force of gravity. Our results suggest the possibility of leveraging the powerful
383 genetic and physiological toolkit of *C. elegans* to elucidate the molecular and circuit
384 mechanisms for gravity sensing – mechanisms that are still elusive.

385

386 **Acknowledgements**

387 We thank Drs. Christopher Fang-Yen and Abraham Wyner for useful discussions. W.L.C was
388 funded by the Ministry of Education of Taiwan, Global Networking Talent 3.0 Plan (GNT3.0),

389 and the Medical Device Innovation Center, National Cheng Kung University.

390

391 **Conflict of Interest**

392 The authors declare no conflict of interest

393

394 **Authors Contributions**

395 DR and HHB planned experiments, HK carried out preliminary experiments, and WLC

396 performed the majority of experiments in the USA and in Taiwan, the latter with supervision

397 by HSC. DR, HHB, and WLC wrote the paper. All authors read and approved the paper,

398

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463 LIST OF FIGURE CAPTIONS

464 1. Experimental set-up (Isometric View)

465 2. **Wild-type animals rotate to align their direction of motion as they descend in solution.**

466 (A-J) 10 video frames spaced 1s apart of a descending young adult, wild-type worm. The
467 red dot indicates the position of the worm's head. The animals are 6-12 mm beneath the
468 water surface. The polar angle varied from 5.1° (frame A) to 141.5° (frame J). (K) The
469 skeletons of the worms from (A-J) were shifted to align their geometric centers to better
470 describe animal's rotation.

471 3. **Wild-type worms change their preferred orientation as they settle in solution.** Kernel-

472 density of wild-type swimmers' orientation angle (θ) at positions 4 mm (Δ , N=145), 12 mm
473 (\circ , N=141), 40 mm (\diamond , N=120), 60 mm (\square , N=133), 80 mm (\blacksquare , N=126), and 100 mm (+,
474 N=123) beneath the liquid surface. The figure was produced with the MatlabTM function
475 "ksdensity" with "bandwidth" of 15. The inset depicts the concentration parameter λ as a
476 function of the animal's position (d mm) beneath the surface. KDEs for depths >100 mm
477 are provided in SI Section S5.

478 4. **Translational velocity and Sedimentation velocity of worms during gravitaxis.**

479 Translational velocity of first day adult WT (N=79) and of motion-impaired adult mutant
480 *unc-54* (N = 52) as functions of $-\cos \theta$, where $\theta = 0$ corresponds to upward orientation.

481 5. **Paralyzed WT worms retain random distribution of their orientation as they settle in**

482 **solution.** Kernel (probability) density estimate (KDE) of heat-shocked paralyzed WT
483 animals at positions 120 (Δ , N=125), 140 (\circ , N=128), 160 (\diamond , N=133), 180 (\square , N=127),
484 and 200 (\blacksquare , N=126), mm beneath the liquid surface. The bandwidth of the KDE smoothing

485 window is 15. See SI for KDEs of paralyzed WT at smaller depths (**Fig. S10**). Inset: the
486 concentration parameter λ as a function of depth. λ remains close to zero consistent with
487 uniform (random) distribution.

488 **6. Wild-type animals rotate to align their direction of motion downward when suspended**
489 **in a solution denser than the animals.** (A-J) 10 video frames spaced 1s apart of a young
490 adult, wild-type worm. The red dot indicates the position of the worm's head. The polar
491 angle varied from 49.1° in frame A to 138.9° in frame J. **(K)** The skeletons of the worms
492 in (A-J) were shifted to align their geometric centers.

493 **7. Gravitaxis does not affected by direction of buoyancy.** The kernel (probability) density
494 estimate of orientation angle of animals suspended in LUDOX HS-40 suspension (density
495 1.1 g/mL and viscosity about 7 times that of water) shortly ($< 2s$) after the animals
496 introduction into the suspension, 5s later, and 10s later. $N_0 = 31$, $N_{5s} = 30$, and $N_{10s} = 36$.
497 In depicting the KDE curves, we used MatlabTM default values.

498 **8. Aging adults have impaired ability to orient with the gravity vector.** Kernel-
499 (probability) density estimate of the orientation angle θ of day 1 (AD 1, solid line) and day
500 6 (AD6, \square) adults at 40 mm beneath liquid surface. The inset depicts the concentration
501 parameter λ as a function of age. For day 6 animals, $p < 0.01$ (**, Mann Whitney Test).
502 $N_{AD1} = 87$, $N_{AD2} = 62$, $N_{AD3} = 60$, $N_{AD4} = 55$, $N_{AD5} = 40$, $N_{AD6} = 50$, In depicting the KDE
503 curves, we used MatlabTM default values.

504 **9. Sensory mutants *che-2* and *osm-6* show defects in downward orientation.** Kernel-
505 density estimate plot of angle of descent of sensory mutants and of wild-type controls at 40
506 mm beneath liquid surface. The distributions of angles of descent of *che-2* and *osm-6*
507 mutants are all broader than that of wild-type animals and approximate random distribution.
508 Compared to WT distribution, $p < 0.0001$ (Mann Whitney Test). $N_{WT}=87$, $N_{che-2}=51$, and
509 $N_{osm-6}=70$. In depicting the KDE curves, we used MatlabTM default values.

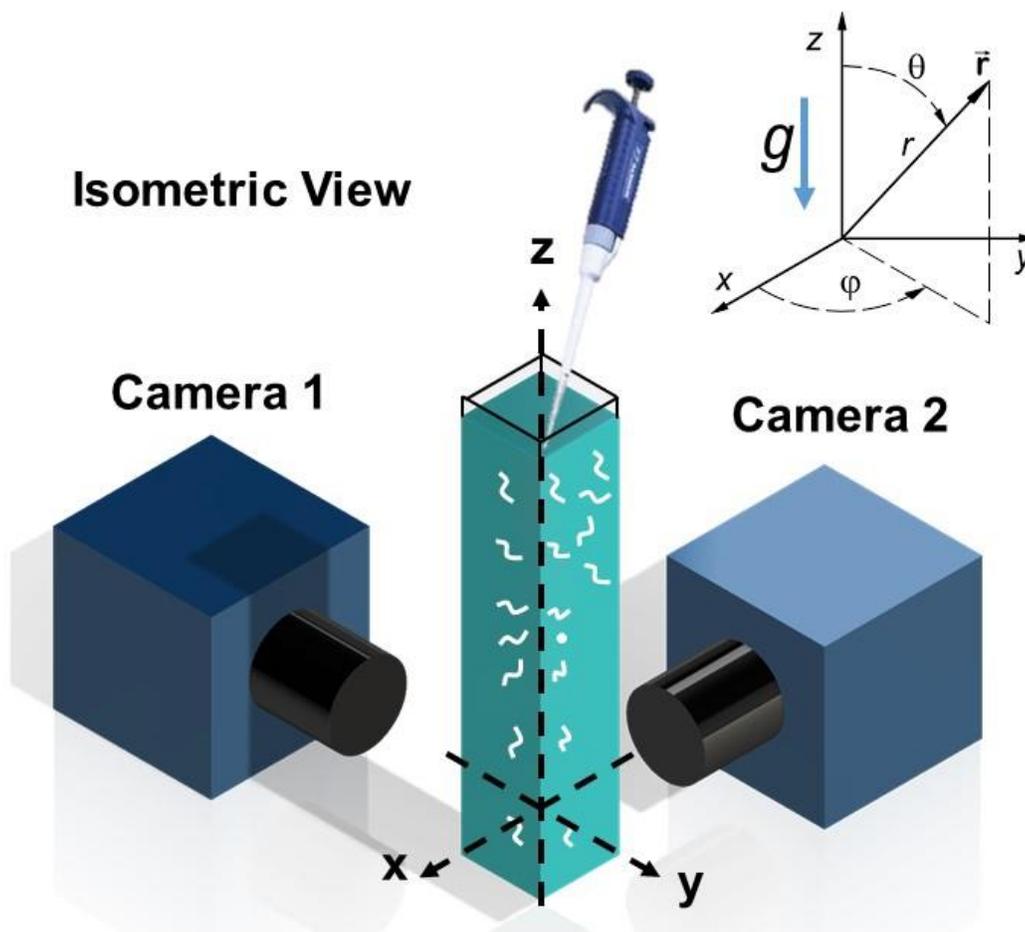


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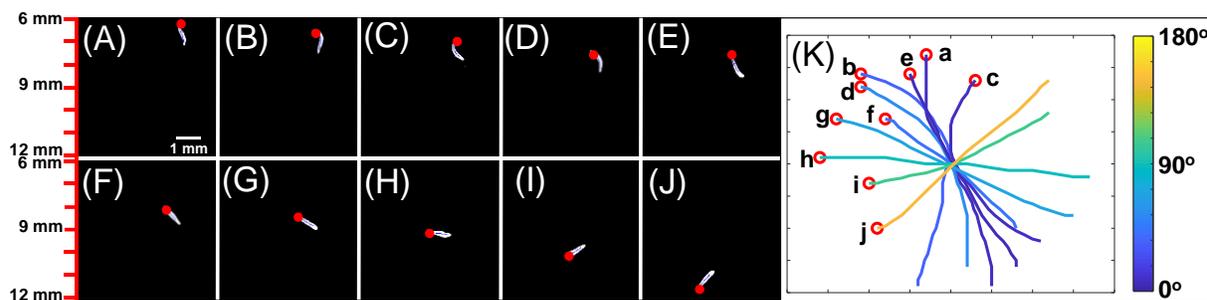


Figure 2: Wild-type animals rotate to align their direction of motion as they descend in solution. (A-J) 10 video frames spaced 1s apart of a descending young adult, wild-type worm. The red dot indicates the position of the worm's head. The animals are 6-12 mm beneath the water surface. The polar angle varied from 5.1° (frame A) to 141.5 (frame J). (K) The skeletons of the worms from (A-J) were shifted to align their geometric centers to better describe animal's rotation.

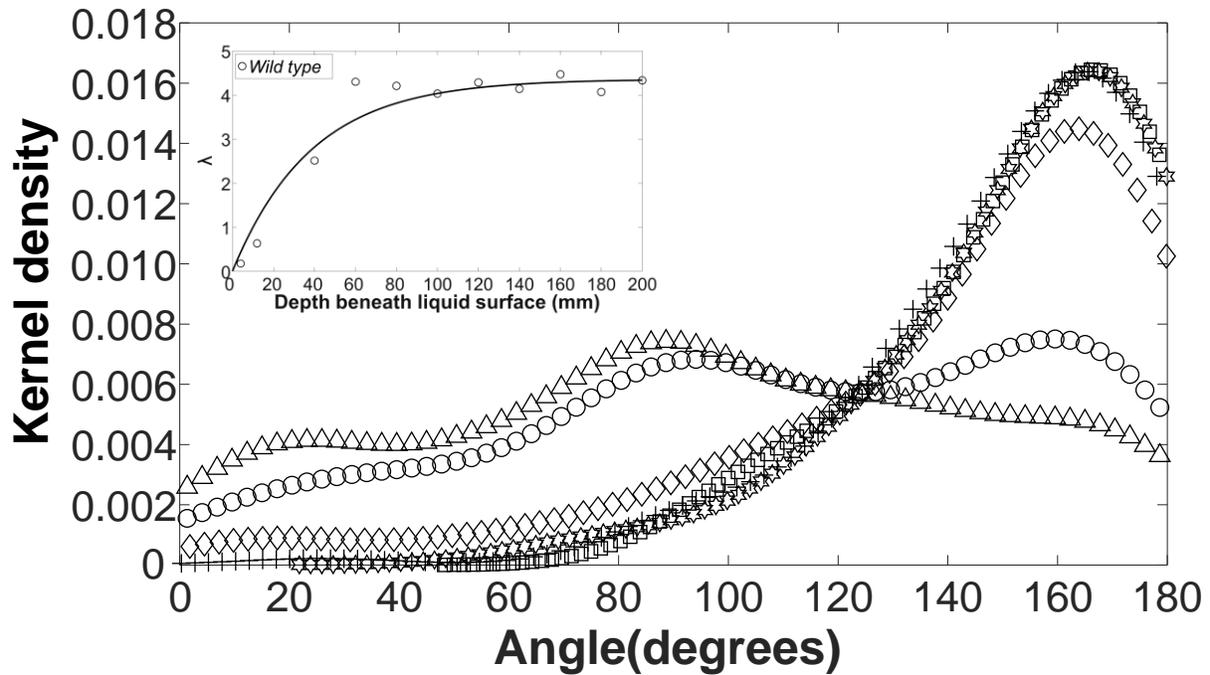


Figure 3: Wild-type worms change their preferred orientation as they settle in solution. Kernel-density of wild-type swimmers' orientation angle (θ) at positions 4 mm (Δ , $N=145$), 12 mm (\circ , $N=141$), 40 mm (\diamond , $N=120$), 60 mm (\square , $N=133$), 80 mm (\star , $N=126$), and 100 mm ($+$, $N=123$) beneath the liquid surface. The figure was produced with the MatlabTM function “ksdensity” with “bandwidth” of 15. The inset depicts the concentration parameter λ as a function of the animal's position (d mm) beneath the surface. KDEs for depths >100 mm are provided in SI Section S5.

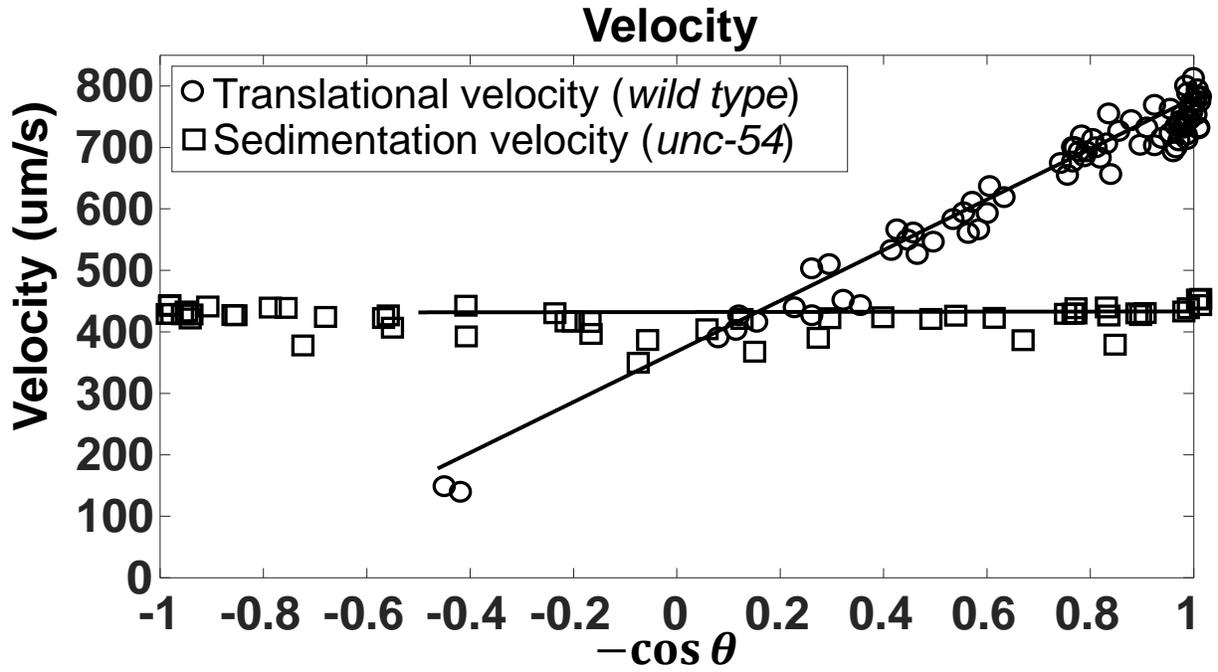


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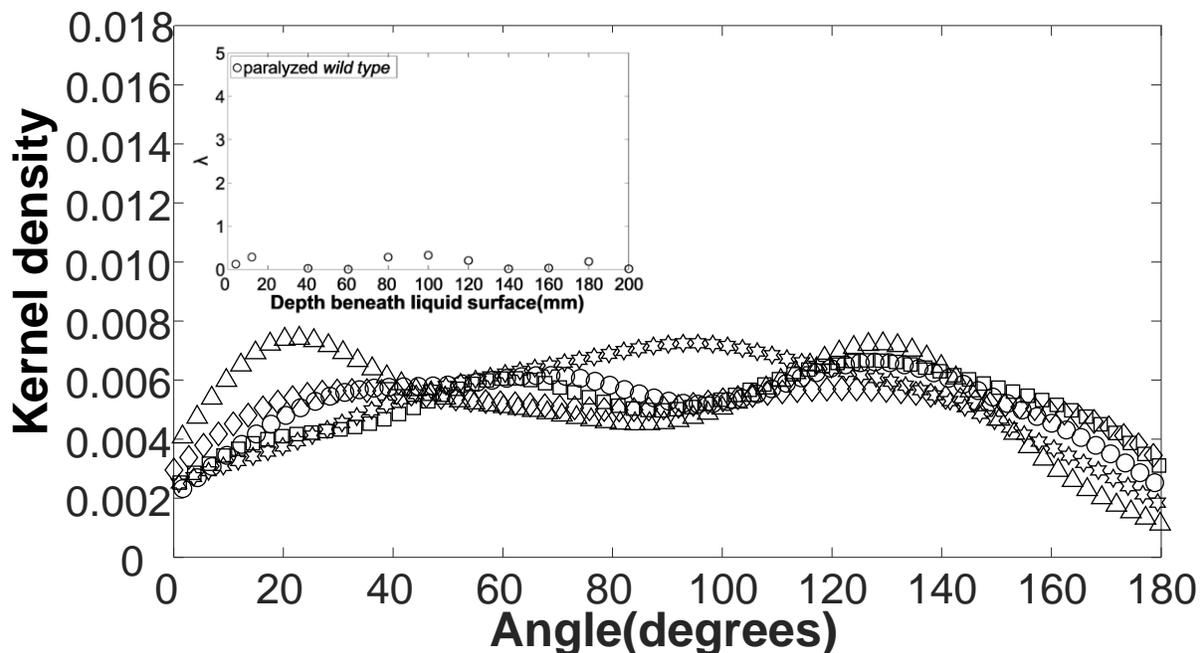


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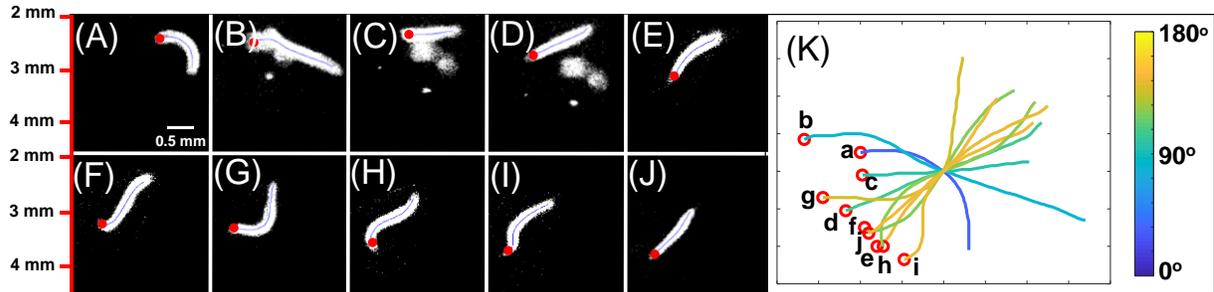


Figure 6: Wild-type animals rotate to align their direction of motion downward when suspended in a solution denser than the animals. (A-J) 10 video frames spaced 1s apart of a young adult, wild-type worm. The red dot indicates the position of the worm's head. The polar angle varied from 49.1° in frame A to 138.9° in frame J. (K) The skeletons of the worms in (A-J) were shifted to align their geometric centers.

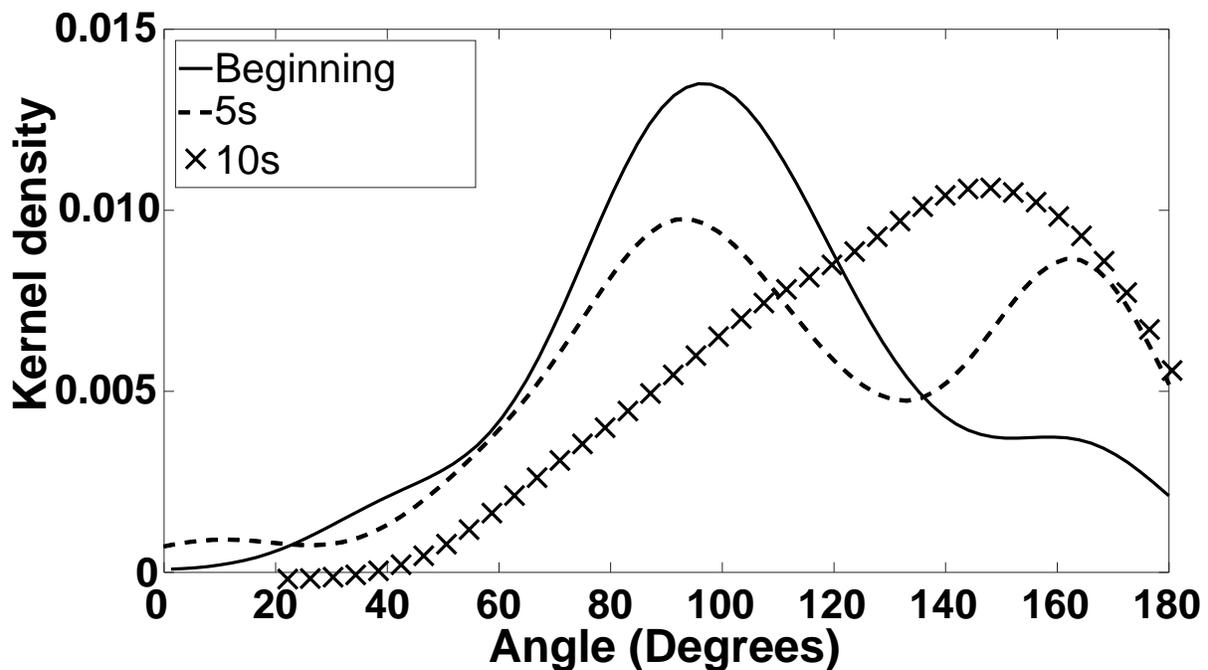


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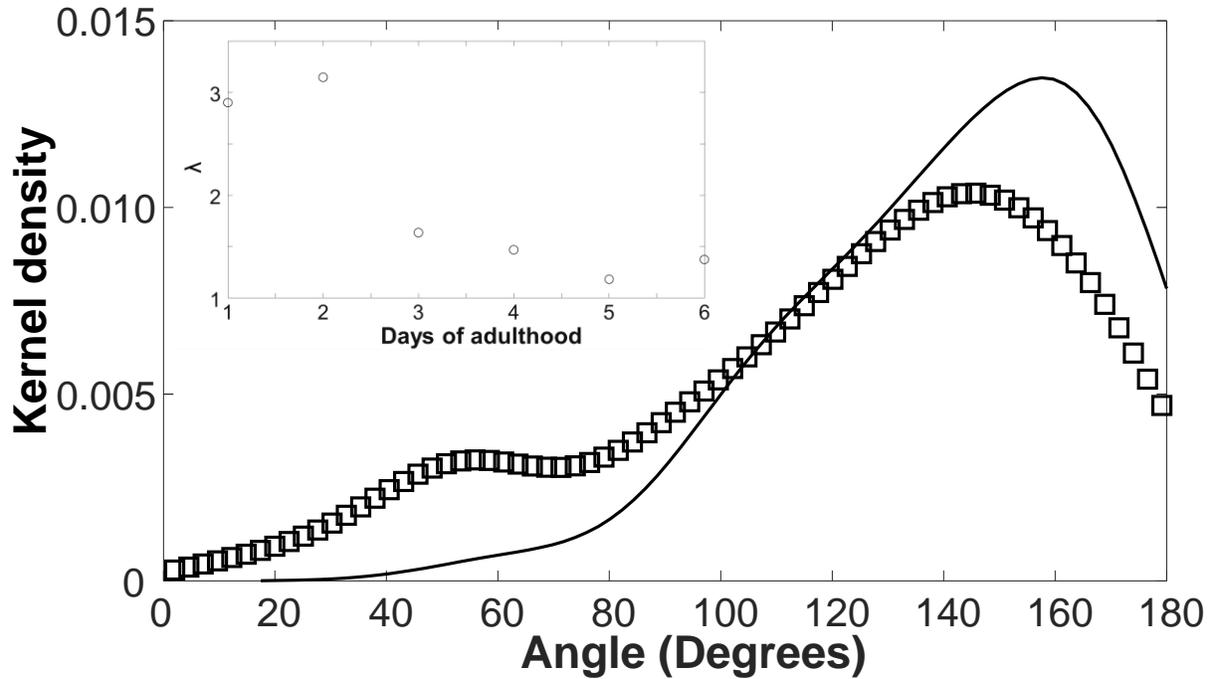


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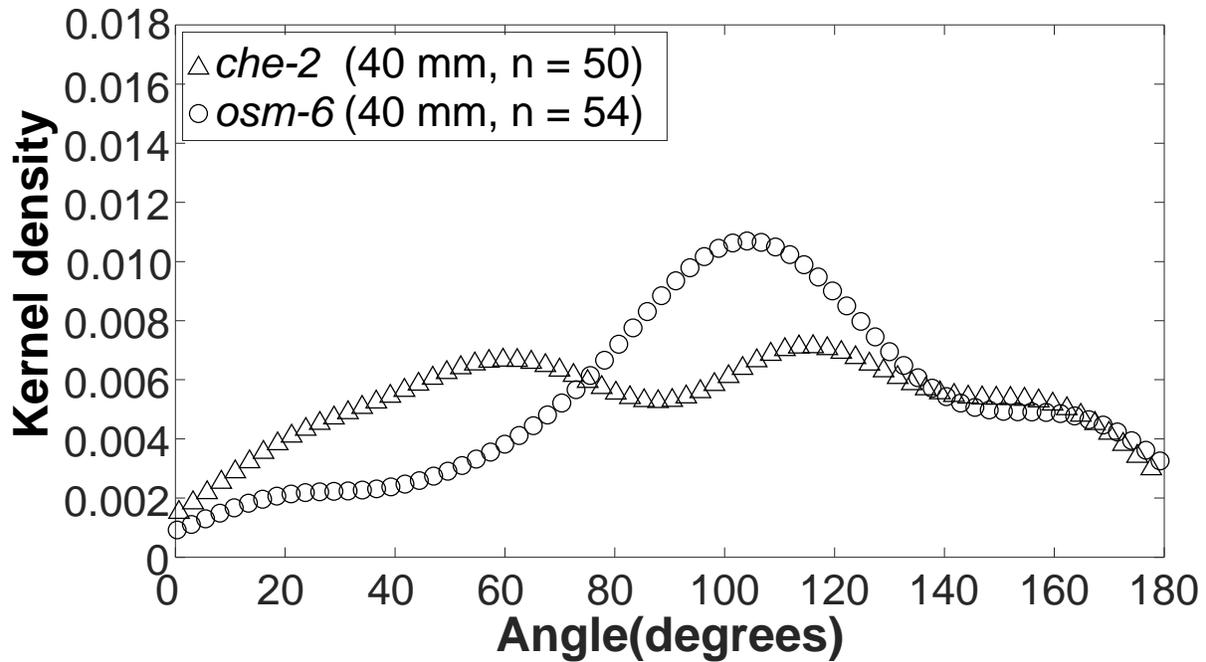


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